

Title: A prospective longitudinal study of retinal structure and function in achromatopsia

Authors:

Jonathan Aboshiha^{1,2}
Adam M. Dubis^{1,2}
Jill Cowing¹
Rachel T.A. Fahy¹
Venki Sundaram²
James W. Bainbridge^{1,2}
Robin R. Ali¹
Alfredo Dubra^{3,4}
Marko Nardini⁶
Andrew R. Webster^{1,2}
Anthony T. Moore^{1,2}
Gary Rubin¹
Joseph Carroll^{3,4,5}
Michel Michaelides^{1,2}

Joseph Carroll and Michel Michaelides are considered joint senior authors.

Corresponding author: Michel Michaelides, UCL Institute of Ophthalmology, 11-43 Bath Street, London, EC1V 9EL, UK. E-mail: michel.michaelides@ucl.ac.uk
Tel. no. 004420 7608 6864

1 UCL Institute of Ophthalmology, University College London, London, UK.

2 Moorfields Eye Hospital, London, UK.

3 Department of Ophthalmology, Medical College of Wisconsin, Milwaukee, Wisconsin.

4 Department of Biophysics, Medical College of Wisconsin, Milwaukee, Wisconsin.

5 Department of Cell Biology, Neurobiology & Anatomy, Medical College of Wisconsin, Milwaukee, Wisconsin.

6 Department of Psychology, Durham University, Durham, UK

Financial Support: Supported by grants from the National Institute for Health Research Biomedical Research Centre at Moorfields Eye Hospital NHS Foundation Trust and UCL Institute of Ophthalmology, Fight for Sight, Moorfields Eye Hospital Special Trustees, The Wellcome Trust [099173/Z/12/Z], Retinitis Pigmentosa Fighting Blindness, and the Foundation Fighting Blindness (USA). Michel Michaelides is supported by a Foundation Fighting Blindness Career Development Award. James W. Bainbridge is an NIHR Research Professor. MCW Funding: NIH grants R01EY017607, P30EY001931, C06RR016511, Foundation Fighting Blindness, and an unrestricted departmental grant from Research to Prevent Blindness (RPB). Alfredo Dubra is the recipient of a Burroughs Wellcome Fund Career Award at the Scientific interface and a Career Development Award from RPB.

Financial Disclosures: No conflicting relationship exists for any author.

Word count: 5058

Running Title: Natural history of achromatopsia

Abstract

Purpose: To longitudinally characterize retinal structure and function in achromatopsia (ACHM) in preparation for clinical gene therapy trials.

Methods: 38 molecularly confirmed ACHM subjects underwent serial assessments, including spectral domain optical coherence tomography (SD-OCT), microperimetry, and fundus autofluorescence (FAF). Foveal structure on SD-OCT was graded and compared for evidence of progression, along with serial measurements of foveal total retinal thickness (FTRT) and outer nuclear layer (ONL) thickness. FAF patterns were characterized and compared over time.

Results: Mean follow-up was 19.5 months (age range at baseline: 6-52 years). Only 2 of 37 subjects (5%) demonstrated change in serial foveal SD-OCT scans. There was no statistically significant change over time in FTRT ($p=0.83$), ONL thickness ($p=0.27$), hyporeflective zone (HRZ) diameter ($p=0.42$), visual acuity ($p=0.89$), contrast sensitivity ($p=0.22$), mean retinal sensitivity ($p=0.84$), and fixation stability ($p=0.58$). Three distinct FAF patterns were observed ($n=30$): central increased FAF ($n=4$), normal FAF ($n=11$), and well-demarcated reduced FAF ($n=15$); with the latter group displaying a slow increase in the area of reduced FAF of 0.03 mm^2 over 19.3 months ($p=0.002$).

Conclusions: Previously published cross-sectional studies have described conflicting findings with respect to the age-dependency of progression. This study, which constitutes the largest and longest prospective longitudinal study of ACHM to date, suggests that although ACHM may be progressive, any such progression is slow and subtle in most patients, and does not correlate with age or genotype. We also describe the first serial assessment of FAF, which is highly variable between individuals, even of similar age and genotype.

Abstract word count: 246

1 INTRODUCTION

2 Achromatopsia (ACHM) is an autosomal recessive cone dysfunction syndrome affecting about 1 in
3 30,000 people, and characterized by the presentation in infancy of pendular nystagmus, poor visual
4 acuity, and photophobia.¹ Electroretinography (ERG) demonstrates absent cone responses and
5 normal or near normal rod responses,^{2,3} with psychophysical testing revealing normal rod function
6 but absent or severely reduced cone function.⁴ To date, five genes have been found to be
7 associated with ACHM, all encoding components of the cone-specific phototransduction cascade.
8 The two most common of these are *CNGA3*⁵ and *CNGB3*,⁶ encoding the α - and β -subunits of the
9 cGMP-gated cation channel respectively, and together account for approximately 70% of cases.⁷
10 Disease-causing sequence variants have also been identified in *GNAT2*,⁸ *PDE6C*,⁹ and *PDE6H*,¹⁰; each
11 responsible for < 2% of cases.⁸⁻¹⁰

12 Several studies have recently demonstrated the effectiveness of using a gene-replacement approach
13 to restore cone function in dog and mouse models of ACHM,¹¹⁻¹⁴ and the neuroprotective protein
14 ciliary neurotrophic factor has also been shown to induce a transient restoration of cone function
15 and visually directed behavior in the *cngb3* dog model.¹⁵ Given these promising results, there are
16 plans to begin human clinical trials in the near future. This makes the accurate measurement and
17 stratification of retinal structure and function in ACHM critical, both in terms of patient selection and
18 subsequent assessment of treatment response.

19 ACHM has been classically described as stationary,^{1,2,7} however, several recent studies have
20 suggested it is a progressive condition. Thiadens et al¹⁶ examined 40 ACHM patients using spectral-
21 domain optical coherence tomography (SD-OCT), and reported that increased cone cell 'decay' and
22 retinal thinning was correlated with age, and began in early childhood. They reported that cone loss,
23 as assessed on SD-OCT, occurred in 42% (8 of 19) of the achromats aged less than 30 years old,
24 whereas this finding was observed in 95% (20 of 21) of patients aged older than 30 years. Thomas et
25 al¹⁷ also noted in their study (n=13) that the existence of a hyporeflective zone (HRZ), and outer
26 nuclear layer (ONL) thinning were both age dependent. In contrast to these studies, Genead et al¹⁸

27 carried out a variety of investigations to assess macular structure, including SD-OCT (n=12), and
28 found that there was significant structural variability even within individuals of the same genotype
29 and age. They also found adaptive optics scanning light ophthalmoscopy (AOSLO) evidence of
30 residual cone inner segment structure in all but one of the nine patients assessed, including the
31 oldest patient (55 years), suggesting that progressive and complete loss of cones in ACHM may not
32 be inevitable, and was not obviously age-dependent. Sundaram et al¹⁹ examined 40 ACHM patients
33 aged 6 to 52 years old, and found no correlation between age and visual acuity, total foveal
34 thickness, foveal ONL thickness, or inner segment ellipsoid (ISe) intensity and cone loss on SD-OCT.

35 All of these studies are inherently limited by their cross-sectional nature, and the debate
36 surrounding whether there is, or is not, age-dependent cone loss highlights the need for prospective
37 longitudinal studies of large cohorts of molecularly-proven patients. Thomas et al²⁰ followed-up
38 eight patients longitudinally, over a mean period of 16 months. They reported that the five younger
39 patients (aged <10 years) demonstrated progressive disturbance at the foveal photoreceptor inner
40 segment/outer segment (IS/OS) junction on OCT; but the three older patients (aged >40 years) did
41 not. Due to the small number of patients they were not able to perform statistical comparisons
42 between the two visits.

43 A further means of assessing macular integrity is through the use of fundus autofluorescence (FAF),²¹
44 which can act as a surrogate measure of photoreceptor health.²² There are limited data concerning
45 FAF patterns in ACHM. Michaelides et al²³ reported that the FAF appearance was normal in five
46 affected members of a *GNAT2* family. In a recent study of 10 achromats, Fahim et al²⁴ suggested an
47 age-dependent change in FAF, with younger patients exhibiting increased foveal autofluorescence
48 (AF), whilst older patients had reduced AF with discrete borders corresponding to outer retinal
49 defects on SD-OCT. Greenberg et al²⁵ also observed FAF abnormalities including increased and/or
50 decreased AF (n=17). However, again these were cross-sectional studies of relatively small size, with
51 no longitudinal FAF data in ACHM published to date. There is a need to assess whether FAF signals in

52 ACHM change over time, which might aid assessment of any progression, genotype characterization,
53 and potentially serve as a marker for response to future interventions.

54 Here we performed serial assessments of visual acuity, contrast sensitivity, microperimetry (MP), SD-
55 OCT, and FAF, in the largest longitudinal study of molecularly proven patients to date, with the goal
56 of providing statistically supported conclusions regarding the relative frequency and rate of any
57 observed progression.

58 **METHODS**

59 *Subjects*

60 Forty subjects that had been characterized both phenotypically (including electrophysiologically) and
61 genotypically in a previously published cross-sectional study¹⁹ were prospectively asked to return for
62 follow-up assessment at an interval of between 12 and 24 months after baseline assessment. The
63 study protocol adhered to the Tenets of the Declaration of Helsinki, and was approved by the
64 Moorfields Eye Hospital Ethics Committee. Informed consent was obtained from all subjects before
65 entering the study.

66 *Clinical assessments*

67 A detailed follow-up assessment was undertaken which included best-corrected visual acuity (BCVA)
68 using an Early Treatment Diabetic Retinopathy Study chart, contrast sensitivity assessment using the
69 Pelli-Robson chart at 1 m, MP, SD-OCT, and FAF. All assessments were prospectively standardized to
70 be undertaken in the same conditions in the follow-up assessment as at baseline assessment.

71 *SD-OCT*

72 After pupillary dilation with tropicamide 1% and phenylephrine 2.5% eye drops, line and volume
73 scans were obtained using a Spectralis SD-OCT on both eyes (Heidelberg Engineering, Heidelberg,
74 Germany) using the same protocol as used in the baseline cross-sectional study.¹⁹ Briefly, the
75 volume acquisition protocol consisted of 49 B-scans (124 μm between scans; 20°x20°), with
76 Automatic Real Time eye tracking used whenever possible. During follow-up assessment, the

77 Spectralis SD-OCT was engaged in its follow-up mode, in order to ensure that the same scanning
78 location was identified at both time points. This is achieved by setting the baseline scan as the
79 reference scan, and the inbuilt software then ensures that the SD-OCT laser is directed to the same
80 retinal location during subsequent image acquisition.²⁶ The lateral scale of each image was
81 estimated using the axial length of the corresponding eye, obtained from the Zeiss IOL Master (Carl
82 Zeiss Meditec, Jena, Germany).

83 Qualitative assessment of foveal morphology was undertaken by grading foveal structure on SD-OCT
84 images into 1 of 5 categories: (1) continuous inner segment ellipsoid (ISe), (2) ISe disruption, (3) ISe
85 absence, (4) presence of an HRZ, or (5) outer retinal atrophy, including loss of RPE (Figure 1). The
86 presence or absence of foveal hypoplasia was also noted, and was defined as the persistence of ≥ 1
87 inner retinal layer (outer plexiform layer, inner nuclear layer, inner plexiform layer, or ganglion cell
88 layer) through the fovea. Consensus grading was established by 3 independent examiners (J.A.,
89 Joe.C., and M.M.). Measurements of foveal total retinal thickness (FTRT) (internal limiting
90 membrane to RPE distance), foveal outer nuclear layer (ONL) thickness and, where relevant, HRZ
91 diameter, were made by a single observer (J.A.) at baseline and follow-up scans, using the digital
92 calipers built-in to the software (Heidelberg Eye Explorer, Heidelberg Engineering, Heidelberg,
93 Germany), and a 1 pixel : 1 μm display with maximal magnification. In cases of foveal hypoplasia,
94 the distance between the posterior outer plexiform layer and the external limiting membrane (ELM)
95 was taken as the ONL thickness. The mean of three measurements was used. Due to the optimum
96 image resolution, imaging speed and follow-up acquisition mode and eye-tracking used in the
97 Spectralis SD-OCT, retinal thickness measurements using this device have been shown to be highly
98 reproducible,²⁶ and this method of assessing foveal thickness in the assessment of macular
99 pathology has been used elsewhere.^{27, 28}

100 ***Fundus autofluorescence***

101 Fundus autofluorescence (FAF) was performed at baseline and follow-up assessments using the AF
102 mode built in to the Spectralis confocal scanning laser ophthalmoscope (Heidelberg Engineering,

103 Heidelberg, Germany). All images were acquired after mydriasis as described above, and after the
104 SD-OCT image acquisition protocol was complete; due to the relatively intense lights used during FAF
105 acquisition and the photophobic nature of achromats. Images were acquired using a 30° field, with
106 an optically pumped solid-state laser producing an excitation wavelength of 488 nm. The induced AF
107 was detected through a barrier filter of 500 nm, and acquired after focusing the retinal image using
108 the 820 nm infrared mode and sensitivity adjustment at 488 nm.

109 In the case of patients who had areas of abnormally reduced AF signal, measurements of this area
110 were performed by describing this region with a mouse-driven cursor, and recording the area given
111 by the in-built image analysis software (Heidelberg Eye Explorer, Heidelberg Engineering, Heidelberg,
112 Germany); this method has been used in the assessment of progressive macular disease
113 elsewhere.^{29,30} All FAF measurements were made by a single observer (J.A.), and for each FAF image
114 the mean of three area measurements was calculated and used for further analysis.

115 ***Microperimetry***

116 Microperimetry (MP) was performed in all subjects on both eyes, and after pupillary dilation, in a
117 darkened room using the MP1 microperimeter (Nidek Technologies, Padova, Italy) in follow-up
118 mode, and otherwise using the same testing conditions as at baseline assessment.¹⁹ Two tests were
119 undertaken on each eye, using a customized test grid of 44 retinal locations situated within an 8°
120 radius in order to cover the macular and para-macular region, and over which the mean retinal
121 sensitivity was recorded. During each test the non-tested contralateral eye was occluded.

122 Background illuminance was set within the mesopic range (1.27cd/m²) and patients maintained
123 fixation by looking at a 2° target. A variable intensity Goldmann size III (4mm²) stimulus of 200ms
124 duration was used as the testing stimulus. A 4-2 testing strategy was employed, with the intensity of
125 the stimulus reduced in 4dB steps until the patient no longer detected it. The stimulus intensity then
126 increased in 2dB steps until detected once again. Projection of the stimulus into the blind spot at 30
127 second intervals tested for false positive errors. Fixation stability was assessed using the bivariate
128 contour ellipse area (BCEA) analysis, which represents an area in degrees where 68% (i.e. one

129 standard deviation) of fixation points are located³¹; this value is reported by the Nidek software. An
130 active eye tracking system ensured accurate stimulus projection in relation to retinal landmarks in
131 order to correct for fixation errors.

132

133 ***Data analysis methods***

134 Histogram plots were used to verify the normality of data before the use of any parametric tests.
135 Statistical analyses were performed using GraphPad Prism, version 5 (GraphPad Software Inc., La
136 Jolla, CA). As the left eye had been selected for further analysis in the original cross-sectional study
137 (which had found no significant difference in measured parameters between eyes),¹⁹ this eye also
138 underwent detailed further analysis in the follow-up study. Patient identification numbers were kept
139 the same as in the cross-sectional study for ease of reference. Variability indicators (\pm) represent one
140 standard deviation in normally distributed metrics.

141 **RESULTS**

142 Thirty-eight of the 40 patients (95%) who were assessed at baseline were able to attend for follow-
143 up assessment within the 12-24 month study recall window. Two female patients were unable to
144 return; one due to the onset of unrelated long-term illness (patient #25) and the other due to
145 pregnancy (patient #32). Of the 38 patients that were followed up, 20 were male (53%) and 18 were
146 female (47%). The mean age of the follow-up cohort was 25 years old at baseline (\pm 12.5; range 6 –
147 52 years), with a mean follow-up interval of 19.5 months (\pm 2.8 months; range 13 – 24 months).
148 Table 1 summarizes the clinical findings, and includes age at baseline, interval of follow-up period,
149 genotype, and functional and structural parameters over time.

150 ***Best corrected visual acuity and contrast sensitivity***

151 The mean BCVA at baseline was 0.92 logarithm of the minimum angle of resolution (logMAR) (\pm
152 0.13; range 0.74 – 1.32 logMAR), and was not significantly different to the mean BCVA at follow-up,
153 which also measured 0.92 logMAR (\pm 0.11; range 0.74 – 1.28 logMAR) (paired t-test; p=0.89). The
154 mean of the logarithm of the contrast sensitivity (logCS) at baseline of 1.16 logCS (\pm 0.23; range 0.5 –

155 1.55 logCS) was also not significantly different to that measured at follow-up of 1.21 logCS (\pm 0.32;
156 range 0.25 – 1.65 logCS) (paired t-test; $p=0.22$); where higher logCS values indicate better contrast
157 sensitivity.

158 ***SD-OCT***

159 In 37 of the 38 patients that underwent serial SD-OCT, foveal scans were acquired at baseline and
160 follow-up that were comparable across time points. In one patient (patient #1) foveal follow-up SD-
161 OCT images could not be obtained due to poor fixation, and this patient was excluded from further
162 SD-OCT analysis. Two of these 37 patients (5%) demonstrated progression on SD-OCT between time
163 points; patient #2 (aged 10 years old at first visit) progressed from category 1 (continuous ISe layer)
164 at baseline to category 2 (ISe disruption) over a 20 month period, and patient #31 (aged 33 years old
165 at first visit) progressed from category 2 (ISe disruption) to category 4 (HRZ) over the same number
166 of months (Figure 2). However, these observed changes were subtle in patient #2, and the SD-OCT
167 findings of patient #31 at baseline, with the high reflectivity foveal lesion, are not typically seen in
168 ACHM. No other patients showed evidence of progression, either in terms of transition to another
169 SD-OCT category or progression within a category (Figure 1).

170 The right eye SD-OCT images of all patients were also graded at baseline and follow-up assessment,
171 given that in the cross-sectional study no parameter measured demonstrated any significant
172 difference between eyes,¹⁹ and so presumably any change over time observed in both eyes would
173 more likely represent real disease progression. Both patients with SD-OCT evidence of progression
174 in the left eye showed similar deterioration in the right eye (Figure 2); whilst the remaining patients
175 had the same SD-OCT appearance in the right eye at baseline and follow-up, as was the case for their
176 left eyes. The changes in patient #2 were again subtle in the right eye, but that the SD-OCT changes
177 were mirrored in both eyes in both patients #2 and #31 would lend support to the view that the
178 morphological change detected in these two patients was real.

179 The remaining 35 patients (95%) showed no evidence of deterioration in SD-OCT appearance on
180 qualitative assessment (Figure 1). No patients had a change in the presence or absence of foveal
181 hypoplasia between baseline and follow-up assessment.

182

183 ***Foveal Total Retinal Thickness, ONL Thickness, and HRZ Diameter***

184 Foveal total retinal thickness (FTRT) was measured in all 37 patients who had serial foveal SD-OCT
185 assessments. There was no statistically significant change between visits (i.e. follow-up thickness
186 minus baseline thickness; negative numbers indicating thinning over time) in FTRT in the cohort,
187 with mean change of $-0.1 \mu\text{m}$ (95% confidence interval (CI) -1.3 to $+1.1 \mu\text{m}$; range -9 to $+4 \mu\text{m}$)
188 (paired t-test; $p=0.83$) (Table 1).

189 Three of the 37 patients (8%) that underwent serial foveal SD-OCT scanning (patients #10, #28 and
190 #30) did not have a sufficiently structurally distinct ONL layer which could be accurately measured,
191 and so were excluded from the ONL thickness analysis. The mean ONL thickness change in the
192 remaining 34 patients of $-0.6 \mu\text{m}$ (95% CI -1.7 to $+0.5 \mu\text{m}$; range -10 to $+3 \mu\text{m}$) was not statistically
193 significant between assessments (paired t-test; $p=0.27$) (Table 1). Only one patient had ONL thinning
194 of $\geq 10 \mu\text{m}$ (patient #2 with $10 \mu\text{m}$); which is in keeping with the qualitative deterioration of outer
195 retinal structure on SD-OCT observed in this patient. In the 9 patients who had a HRZ at both time
196 point assessments, there was no statistically significant change in the diameter of the HRZ over a
197 mean follow-up time of 19.6 months in this group (paired t-test; $p=0.42$).

198

199 ***Microperimetry***

200 All 38 patients underwent follow-up MP testing on ≥ 2 occasions, and the mean of these tests was
201 used for subsequent analysis. The mean retinal sensitivity of this cohort at baseline was $16.6 \text{ dB} (\pm$
202 3.4 ; range $3.1 - 19.9 \text{ dB}$), and was not significantly different to that measured at follow-up ($16.5 \text{ dB} \pm$
203 3.07 ; range $5.8 - 16.5 \text{ dB}$) (paired t-test; $p=0.84$) (Table 1). The fixation stability measured at baseline

204 (median = 9.1°; range 1.7 – 65°) was also not significantly different from that at follow-up (median =
205 8.2°; range 1.7 – 61.8°) (Wilcoxon matched-pairs signed rank test; $p = 0.58$) (Table 1). At baseline
206 assessment six subjects had a scotoma (defined as 0 dB sensitivity in ≥ 1 retinal location). Follow-up
207 MP assessment also showed that the same 6 patients had scotomas. In addition, one further patient
208 (patient # 5, aged 17 years old at baseline assessment) developed a scotoma that was not apparent
209 on initial testing, and which was evident in both follow-up MP tests 16 months later (Figure 3).
210 However, the change in point sensitivity is small and would fall within inter-test variability limits, as
211 can be seen in the variation in sensitivities in the points around those that developed an absolute
212 scotoma.

213

214 ***Fundus autofluorescence***

215 In 30 of the 38 patients (84%), baseline and follow-up FAF images were of sufficient quality to enable
216 accurate classification and, where necessary, measurement of the area of reduced FAF signal.
217 Several patients experienced discomfort with the bright light needed to acquire FAF images, and
218 given their extreme photophobia, this contributed to relatively lower patient concordance and
219 successful acquisition compared to other serial assessment modalities. We observed three FAF
220 patterns at baseline ($n=30$): 1) a normal FAF pattern - seen in 11 patients (37%); 2) an abnormal
221 central increase in FAF – seen in 4 patients (13%); and 3) a discretely bordered abnormal central
222 reduction in FAF – seen in 15 patients (50%) (Figure 4). No patients changed the type of FAF pattern
223 they displayed between baseline and follow-up assessments, and no patients had different FAF
224 patterns between eyes.

225 Given that the reduced FAF pattern had well-demarcated borders amenable to quantification, the
226 area of reduced FAF was measured at baseline and follow-up in the 15 patients with this FAF
227 phenotype. There was a small but statistically significant increase in median area of reduced FAF
228 between assessments (Wilcoxon matched-pairs test; $p=0.002$), with the median change being +0.03

229 mm² (IQR +0.01 to +0.13 mm²; range 0 to +0.28 mm²), where positive numbers indicate an increase
230 in the area of reduced FAF signal over the mean follow-up time in this sub-group of 19.3 months.

231 The mean age of patients in the group with a normal FAF appearance was 27.6 years (range 10 - 52
232 years); 16.0 years (range 11 - 29 years) in the group with an increased FAF pattern and 29.3 years
233 (range 11 - 49 years) in the group with a reduced FAF pattern. There was no statistically significant
234 difference in the ages between the three FAF pattern groups (Kruskal Wallis test; p<0.05). In one of
235 the two patients in whom we detected evidence of progressive change on SD-OCT (patient #31),
236 there was only a minimal increase in the area of reduced FAF area of +0.01 mm² over a follow-up
237 time of 20 months; the other patient who progressed on SD-OCT (patient #2) had a normal FAF
238 pattern at both time points.

239 We observed a statistically significant correlation between more disordered SD-OCT structure
240 (category 1 to 5) and a more abnormal FAF pattern (graded from normal through central increased
241 FAF to central decreased FAF) (Spearman r = 0.55; p=0.002).

242

243 ***Genotype-phenotype correlation***

244 Of the two patients that progressed on qualitative SD-OCT assessment, one had disease-causing
245 variants in *CNGA3*, and the other in *CNGB3*, suggesting no significant association of progression in
246 *CNGA3* vs. *CNGB3*. This would also be in agreement with our previous findings that there was no
247 association between severity of SD-OCT phenotype or presence of foveal hypoplasia, and
248 genotype.¹⁹ However, with only two patients appearing to show progression on SD-OCT, the
249 numbers of progressors by genotype are too small to be anything other than suggestive.

250 When we analyzed the *CNGA3* and *CNGB3* groups separately, we again did not find any statistically
251 significant difference between the baseline and follow-up measures in BCVA (Wilcoxon matched-
252 pairs test; *CNGA3* p=0.75; *CNGB3* p=0.72), contrast sensitivity (*CNGA3* p=0.06; *CNGB3* p=0.11), MP
253 mean sensitivity (*CNGA3* p=0.33; *CNGB3* p=1.0) or MP mean BCEA (*CNGA3* p=1.0; *CNGB3* p=0.41).

254 There was also no statistically significant difference between the *CNGA3* and *CNGB3* groups in the
255 change in FTRT or ONL thickness (FTRT: Mann Whitney U test; p=0.33; ONL thickness: unpaired t-
256 test; p=0.58).

257 In the case of the *CNGA3* and *CNGB3* patients, there was no significant association between the
258 numbers of patients exhibiting each of the three observed FAF patterns and their genotype (Chi²
259 test; p=0.40).

260

261 **DISCUSSION**

262 This study is, to the best of our knowledge (PubMed search 06/02/2014; keywords: achromatopsia,
263 rod monochromatism), the largest prospective longitudinal study of ACHM with the longest mean
264 follow-up time of 19.5 months, and sufficient patient numbers for statistical analysis looking for
265 change over time.

266 Our cohort did not show any evidence of a statistically significant change over time in BCVA, contrast
267 sensitivity, or MP measures of retinal sensitivity and fixation stability, either as whole, or as separate
268 *CNGA3* and *CNGB3* genotype sub-groups. Only two of 37 patients (5%) followed up longitudinally
269 demonstrated evidence of progressive outer retinal changes according to independently graded
270 qualitative SD-OCT analysis over the follow-up period. Thirty-five patients (95%) demonstrated no
271 qualitative changes on their SD-OCT images between baseline and follow-up assessment. This agrees
272 with our longitudinal quantitative assessment of both TRFT and ONL thickness, both of which
273 showed no statistically significant change in the cohort over the time course of this study. Neither of
274 the two patients (#2 and #31) that had a qualitative change in SD-OCT had any significant
275 deterioration over time (defined as worsening of more than two standard deviations from baseline
276 measurements) in their BCVA, contrast sensitivity, MP mean sensitivity or MP mean BCEA. It has
277 been shown in other inherited retinal conditions that SD-OCT changes may precede deterioration of
278 clinical assessment parameters such as BCVA³² and FAF³³, and so despite a failure to demonstrate

279 deterioration in other clinical measures used in this study, the SD-OCT changes in these two patients
280 likely represent real change. Serial assessment using other sensitive retinal imaging techniques such
281 as AOSLO, may shed further light on the earliest indicators of any progression in ACHM.

282 We saw no patients with an ONL or central macular thickness (CMT) thinning of $>10\ \mu\text{m}$, which
283 contrasts with Thomas et al²⁰, who report that the five pediatric patients in their study (aged <10
284 years at first visit) showed SD-OCT evidence of thinning of both CMT and ONL over a mean follow-up
285 time of 13 months. Three of these 5 children appear to have had an ONL thinning of $>10\ \mu\text{m}$,
286 although the numerical values are not stated. The three adults in their study (aged >40 years)
287 showed only minimal variation in these parameters over a mean follow-up of 20 months, in keeping
288 with our study. However, in at least two of the children, the evidence for progression was not
289 unequivocal. Thomas et al²⁰ described how patients #1 and #2 had developed an outer retinal
290 hyper-reflective zone, which they interpreted as an indication of progression. It should be noted
291 however, that the same two patients had a generalized increase in reflectivity of several other layers
292 in their respective follow-up OCT images, including the nerve fiber layer and ganglion cell layer, and
293 so the possibility that this apparent increase in outer retinal reflectivity over time may be artifactual
294 remains. Furthermore, they were unable to perform any statistical analyses due to their small
295 sample size. The reason for the difference in our respective findings regarding the evidence for
296 progressive thinning of the ONL and/or CMT may be related to the fact that we had only 3 children
297 aged ≤ 10 years old at first visit. However, our cohort did contain a total of eight children aged ≤ 12
298 years old at first visit, and the much larger number of patients in our cohort and the consequent
299 ability to carry out a statistical analysis would lend further support to our findings. We also observed
300 no statistically significant change in the diameter of the HRZ in the 9 patients in our study who had
301 this finding at both time point assessments.

302

303 ***Fundus Autofluorescence***

304 Our findings of three distinct FAF patterns is in broad agreement with those of Fahim et al²⁴. We also
305 found a very small but statistically significant increase in the area of reduced FAF over time amongst
306 patients that displayed that pattern; indicating that serial FAF may play a role in assessing change in
307 ACHM. There was no statistically significant difference between ages in the three FAF patterns.
308 However, no patients changed their FAF pattern during the time-course of this study, and the rate of
309 increase in the size of reduced FAF was very slow (median 0.02 mm²/year); by comparison, FAF
310 atrophy rates in age-related macular degeneration (AMD) are several orders of magnitude greater,
311 in the realm of several mm² / year.²⁹ In addition, there were three patients aged ≤ 13 years in the
312 group that had a normal FAF pattern, and one young patient (aged 11 years old) that had a reduced
313 FAF pattern. This again is in keeping with a highly variable disease, where age and genotype are
314 insufficient predictors of phenotype (Figure 4). However, caution needs to be exercised in over-
315 interpreting this data and no definitive conclusions can be reached without larger cohorts.

316 We also observed a moderately strong correlation between degree of foveal SD-OCT structural
317 disorganization and degree of FAF abnormality. FAF intensity may plausibly progress in sequence,
318 from normal signal through increased FAF signal to subsequent FAF signal reduction. It is also
319 possible that the five SD-OCT categories represent a sequence in the severity of foveal outer retinal
320 structure disorganization (from a continuous ISe to outer retinal atrophy); thereby suggesting that
321 the observed correlation of SD-OCT outer retinal architecture to FAF pattern may be of prognostic
322 value, in addition to potentially acting as a further possible metric in treatment trials. Longer follow-
323 up with larger molecularly confirmed cohorts, combined with more quantitative assessment of FAF,
324 are required to establish whether SD-OCT structural abnormality precedes or follows FAF
325 abnormalities in ACHM.

326 The diagnostic and prognostic relevance of FAF in ACHM has yet to be established. Robson et al³⁴
327 demonstrated that increased FAF in cone or cone-rod dystrophy is associated with reduced rod and
328 cone sensitivity, with scotopic sensitivity reductions being milder than photopic losses. In contrast, in

329 AMD increased FAF may be more indicative of rod as opposed to cone dysfunction.³⁵ We found that
330 the MP mean retinal sensitivity (averaged between the two assessments at each time point) was
331 significantly higher in the increased FAF group (median 18.71 dB) compared to the decreased FAF
332 group (median 17.33 dB) (Mann Whitney U test; p=0.008), but did not find any statistically
333 significant difference in retinal sensitivity between normal FAF and increased FAF groups (Mann
334 Whitney U test; p=0.19). In terms of monitoring for progressive change, it would seem that this
335 measure, like the others we have investigated, would need to be assessed over a greater time span,
336 given the likely slow rate of change – and again given the variability, this highlights the need to
337 consider patients on an individual basis in terms of potential suitability for intervention. The use of
338 further quantitative FAF analyses^{24, 35, 36} may increase the sensitivity of FAF assessment in ACHM, but
339 it is still likely that any progression by this measure will occur at a very slow rate in a subset of
340 patients.

341

342 **Age-dependency of progression**

343 In their retrospective study, Thiadens et al³⁷ reported that, over a mean follow-up of 15 years, 12%
344 of the ACHM patients they reviewed had a deterioration in BCVA; in contrast with other cross-
345 sectional studies which have not found an age-dependent deterioration in BCVA.^{16, 17, 19} In their
346 longitudinal study of 8 patients, Thomas et al²⁰ also showed no evidence of a decrease in BCVA over
347 the mean follow-up of 18 months. Khan et al³ also reported that 6 to 12 years had elapsed between
348 their detecting and subsequently failing to detect photopic ERG response in two affected adult
349 *CNGB3* individuals. Given the aforementioned timespans, it is likely that any progression in this
350 condition is very slow and possibly subtle; it may be that to see clear evidence of progression in
351 more patients, longitudinal studies of a much longer time course that may extend into many years or
352 even decades would be required.

353 There is also significant phenotypic variation between individuals in terms of the existence and time
354 course of any such progression. This phenotypic variability is illustrated in the findings of OCT
355 studies^{16, 19, 38-40} and AOSLO studies.^{18, 41, 42} Such variation may confound attempts to look for
356 progression en bloc in what is likely to be a more heterogeneous condition than previously thought.
357 Our data suggests that each individual patient's phenotype, and the significance and time course of
358 any progression, does not correlate with genotype and is highly variable between individuals.
359 Probing this more deeply may require more detailed longitudinal cellular phenotyping with
360 ultrastructural assessments such as AOSLO, potentially in concert with AO-guided psychophysical
361 assessments. The need for such deep-phenotyping has clear implications for the selection and
362 monitoring of patients for anticipated therapeutic interventions. For most of the retinal parameters
363 measured, our initial cross-sectional study did not find any association with *CNGA3* or *CNGB3*
364 genotype.¹⁹ However, we did observe that retinal sensitivity was significantly higher in the *CNGB3*
365 group. This longitudinal study did not identify any measured parameters that worsened significantly
366 between time points when each genotype group was analyzed alone.

367 The longitudinal findings herein broadly agree with our initial cross-sectional study that age and
368 genotype are not necessarily the critical factors in selecting patients for gene therapy. The vast
369 majority of parameters assessed in this study did not change over time, and those that did, either
370 did so very slowly, or in a very small proportion of patients. This supports the view that the potential
371 treatment window for gene therapy may not only be wider in terms of age than has been previously
372 suggested, but that it may remain open for a longer period. Our data also lend support to the view
373 that ACHM is heterogeneous, and that factors other than age and genotype are likely to influence
374 the phenotype. Such factors may have an as of yet unknown influence on any planned therapeutic
375 interventions.

REFERENCES

1. Michaelides M, Hunt DM, Moore AT. The cone dysfunction syndromes. *Br J Ophthalmol*. 2004;88(2):291-7. Epub 2004/01/23. doi: 10.1136/bjo.2003.027102. PubMed PMID: 14736794; PubMed Central PMCID: PMC1771989.
2. Andréasson S, Tornqvist K. Electroretinograms in patients with achromatopsia. *Acta Ophthalmologica*. 1991;69(6):711-6.
3. Khan NW, Wissinger B, Kohl S, Sieving PA. CNGB3 achromatopsia with progressive loss of residual cone function and impaired rod-mediated function. *Investigative ophthalmology & visual science*. 2007;48(8):3864-71.
4. Hess R, Nordby K. Spatial and temporal limits of vision in the achromat. *The Journal of physiology*. 1986;371(1):365-85.
5. Wissinger B, Jägle H, Kohl S, Broghammer M, Baumann B, Hanna DB, et al. Human rod monochromacy: linkage analysis and mapping of a cone photoreceptor expressed candidate gene on chromosome 2q11. *Genomics*. 1998;51(3):325-31.
6. Sundin OH, Yang J-M, Li Y, Zhu D, Hurd JN, Mitchell TN, et al. Genetic basis of total colourblindness among the Pingelapese islanders. *Nature genetics*. 2000;25(3):289-93.
7. Johnson S. Achromatopsia caused by novel mutations in both CNGA3 and CNGB3. *Journal of Medical Genetics*. 2004;41(2):20e-. doi: 10.1136/jmg.2003.011437.
8. Kohl, Baumann B, Rosenberg T, Kellner U, Lorenz B, Vadalà M, et al. Mutations in the Cone Photoreceptor G-Protein α -Subunit Gene GNAT2 in Patients with Achromatopsia. *The American Journal of Human Genetics*. 2002;71(2):422-5. doi: <http://dx.doi.org/10.1086/341835>.
9. Chang B, Grau T, Dangel S, Hurd R, Jurklics B, Sener EC, et al. A homologous genetic basis of the murine cpfl1 mutant and human achromatopsia linked to mutations in the PDE6C gene. *Proceedings of the National Academy of Sciences*. 2009;106(46):19581-6.
10. Kohl S, Coppieters F, Meire F, Schaich S, Roosing S, Brennenstuhl C, et al. A Nonsense Mutation in PDE6H Causes Autosomal-Recessive Incomplete Achromatopsia. *The American Journal of Human Genetics*. 2012;91(3):527-32. doi: <http://dx.doi.org/10.1016/j.ajhg.2012.07.006>.
11. Alexander JJ, Umino Y, Everhart D, Chang B, Min SH, Li Q, et al. Restoration of cone vision in a mouse model of achromatopsia. *Nature medicine*. 2007;13(6):685-7.
12. Komaromy AM, Alexander JJ, Rowlan JS, Garcia MM, Chiodo VA, Kaya A, et al. Gene therapy rescues cone function in congenital achromatopsia. *Hum Mol Genet*. 2010;19(13):2581-93. Epub 2010/04/10. doi: 10.1093/hmg/ddq136. PubMed PMID: 20378608; PubMed Central PMCID: PMC2883338.
13. Michalakis S, Mühlfriedel R, Tanimoto N, Krishnamoorthy V, Koch S, Fischer MD, et al. Restoration of cone vision in the CNGA3^{-/-} mouse model of congenital complete lack of cone photoreceptor function. *Molecular Therapy*. 2010;18(12):2057-63.
14. Carvalho LS, Xu J, Pearson RA, Smith AJ, Bainbridge JW, Morris LM, et al. Long-term and age-dependent restoration of visual function in a mouse model of CNGB3-associated achromatopsia following gene therapy. *Hum Mol Genet*. 2011;20(16):3161-75. Epub 2011/05/18. doi: 10.1093/hmg/ddr218. PubMed PMID: 21576125; PubMed Central PMCID: PMC3140821.
15. Wen R, Tao W, Li Y, Sieving PA. CNTF and retina. *Progress in retinal and eye research*. 2012;31(2):136-51.
16. Thiadens AA, Somervuo V, van den Born LI, Roosing S, van Schooneveld MJ, Kuijpers R, et al. Progressive loss of cones in achromatopsia: an imaging study using spectral-domain optical coherence tomography. *Investigative ophthalmology & visual science*. 2010;51(11):5952-7.

17. Thomas MG, Kumar A, Kohl S, Proudlock FA, Gottlob I. High-resolution in vivo imaging in achromatopsia. *Ophthalmology*. 2011;118(5):882-7.
18. Genead MA, Fishman GA, Rha J, Dubis AM, Bonci DMO, Dubra A, et al. Photoreceptor structure and function in patients with congenital achromatopsia. *Investigative ophthalmology & visual science*. 2011;52(10):7298-308.
19. Sundaram V, Wilde C, Aboshiha J, Cowing J, Han C, Langlo CS, et al. Retinal Structure and Function in Achromatopsia: Implications for Gene Therapy. *Ophthalmology*. 2013(0). doi: 10.1016/j.ophtha.2013.08.017. PubMed PMID: 24148654.
20. Thomas MG, McLean RJ, Kohl S, Sheth V, Gottlob I. Early signs of longitudinal progressive cone photoreceptor degeneration in achromatopsia. *British Journal of Ophthalmology*. 2012;96(9):1232-6.
21. Delori FC, Dorey CK, Staurenghi G, Arend O, Goger DG, Weiter JJ. In vivo fluorescence of the ocular fundus exhibits retinal pigment epithelium lipofuscin characteristics. *Investigative ophthalmology & visual science*. 1995;36(3):718-29.
22. Schmitz-Valckenberg S, Holz FG, Bird AC, Spaide RF. Fundus autofluorescence imaging: review and perspectives. *Retina*. 2008;28(3):385-409.
23. Michaelides M, Aligianis I, Holder G, Simunovic M, Mollon J, Maher E, et al. Cone dystrophy phenotype associated with a frameshift mutation (M280fsX291) in the α -subunit of cone specific transducin (GNAT2). *British journal of ophthalmology*. 2003;87(11):1317-20.
24. Fahim AT, Khan NW, Zahid S, Schachar IH, Branham K, Kohl S, et al. Diagnostic Fundus Autofluorescence Patterns in Achromatopsia. *American journal of ophthalmology*. 2013.
25. Greenberg JP, Sherman J, Zweifel SA, Chen RW, Duncker T, Kohl S, et al. Spectral-Domain Optical Coherence Tomography Staging and Autofluorescence Imaging in Achromatopsia. *JAMA Ophthalmol*. 2014. Epub 2014/02/08. doi: 10.1001/jamaophthalmol.2013.7987. PubMed PMID: 24504161.
26. Menke MN, Dabov S, Knecht P, Sturm V. Reproducibility of retinal thickness measurements in healthy subjects using spectralis optical coherence tomography. *American journal of ophthalmology*. 2009;147(3):467-72.
27. Maheshwary AS, Oster SF, Yuson RMS, Cheng L, Mojana F, Freeman WR. The Association Between Percent Disruption of the Photoreceptor Inner Segment–Outer Segment Junction and Visual Acuity in Diabetic Macular Edema. *American Journal of Ophthalmology*. 2010;150(1):63-7.e1. doi: <http://dx.doi.org/10.1016/j.ajo.2010.01.039>.
28. Oster SF, Mojana F, Brar M, YUSON RM, Cheng L, Freeman WR. Disruption of the photoreceptor inner segment/outer segment layer on spectral domain-optical coherence tomography is a predictor of poor visual acuity in patients with epiretinal membranes. *Retina*. 2010;30(5):713-8.
29. Holz FG, Bindewald-Wittich A, Fleckenstein M, Dreyhaupt J, Scholl HP, Schmitz-Valckenberg S. Progression of geographic atrophy and impact of fundus autofluorescence patterns in age-related macular degeneration. *American journal of ophthalmology*. 2007;143(3):463-72. e2.
30. Dreyhaupt J, Mansmann U, Pritsch M, Dolar-Szczasny J, Bindewald A, Holz F. Modelling the natural history of geographic atrophy in patients with age-related macular degeneration. *Ophthalmic epidemiology*. 2005;12(6):353-62.
31. Crossland MD, Dunbar HM, Rubin GS. Fixation stability measurement using the MP1 microperimeter. *Retina*. 2009;29(5):651-6.
32. Park SJ, Woo SJ, Park KH, Hwang J-M, Chung H. Morphologic photoreceptor abnormality in occult macular dystrophy on spectral-domain optical coherence tomography. *Investigative Ophthalmology & Visual Science*. 2010;51(7):3673-9.
33. Gomes NL, Greenstein VC, Carlson JN, Tsang SH, Smith RT, Carr RE, et al. A comparison of fundus autofluorescence and retinal structure in patients with Stargardt disease. *Investigative ophthalmology & visual science*. 2009;50(8):3953-9.
34. Robson AG, Michaelides M, Luong V, Holder GE, Bird AC, Webster AR, et al. Functional correlates of fundus autofluorescence abnormalities in patients with RPGR or RIMS1 mutations causing cone or cone–rod dystrophy. *British Journal of Ophthalmology*. 2008;92(1):95-102.

35. Scholl HP, Bellmann C, Dandekar SS, Bird AC, Fitzke FW. Photopic and scotopic fine matrix mapping of retinal areas of increased fundus autofluorescence in patients with age-related maculopathy. *Investigative ophthalmology & visual science*. 2004;45(2):574-83.
36. Smith RT, Koniarek JP, Chan J, Nagasaki T, Sparrow JR, Langton K. Autofluorescence characteristics of normal foveas and reconstruction of foveal autofluorescence from limited data subsets. *Investigative ophthalmology & visual science*. 2005;46(8):2940-6.
37. Thiadens AA, Slingerland NW, Roosing S, van Schooneveld MJ, van Lith-Verhoeven JJ, van Moll-Ramirez N, et al. Genetic etiology and clinical consequences of complete and incomplete achromatopsia. *Ophthalmology*. 2009;116(10):1984-9. e1.
38. Varsányi B, Somfai GM, Lesch B, Vámos R, Farkas Á. Optical coherence tomography of the macula in congenital achromatopsia. *Investigative ophthalmology & visual science*. 2007;48(5):2249-53.
39. Barthelmes D, Sutter FK, Kurz-Levin MM, Bosch MM, Helbig H, Niemeyer G, et al. Quantitative analysis of OCT characteristics in patients with achromatopsia and blue-cone monochromatism. *Investigative ophthalmology & visual science*. 2006;47(3):1161-6.
40. Nishiguchi KM, Sandberg MA, Gorji N, Berson EL, Dryja TP. Cone cGMP-gated channel mutations and clinical findings in patients with achromatopsia, macular degeneration, and other hereditary cone diseases. *Human mutation*. 2005;25(3):248-58.
41. Merino D, Duncan JL, Tiruveedhula P, Roorda A. Observation of cone and rod photoreceptors in normal subjects and patients using a new generation adaptive optics scanning laser ophthalmoscope. *Biomedical optics express*. 2011;2(8):2189-201.
42. Carroll J, Choi SS, Williams DR. In vivo imaging of the photoreceptor mosaic of a rod monochromat. *Vision research*. 2008;48(26):2564-8. doi: <http://dx.doi.org/10.1016/j.visres.2008.04.006>.

Acknowledgements

The authors thank Vincent Rocco and Monica Clemo for their work on SD-OCT image acquisition, and all the patients and their families for their time and cooperation in taking part in this study.

Figure Captions

FIGURE 1 Longitudinal spectral-domain optical coherence tomography (SD-OCT) scans in achromatopsia. Left panel column: Baseline scans indicating genotype, patient number and age at initial scan. Right panel column: Follow-up scans indicating patient number and interval between baseline and follow-up scans in months. Central panel column shows foveal magnification (x3) of corresponding scan (grey arrows). Shown are the left eyes of a typical subject from each of the five

SD-OCT categories, all of whom demonstrated no change in SD-OCT structure over the study period (from the top row down: patient #35 with a continuous inner segment ellipsoid (ISe) band at both time points; patient #21 with a disrupted ISe layer at both time points; patient #11 with an absent ISe layer at both time points; patient #12 with a hyporeflective zone (HRZ) at both time points; and patient #30 with outer retinal atrophy at both time points). Non-magnified images represent 4500 μ m horizontally and 1000 μ m vertically; image scaling has been corrected for axial length. Scale bar, 200 μ m.

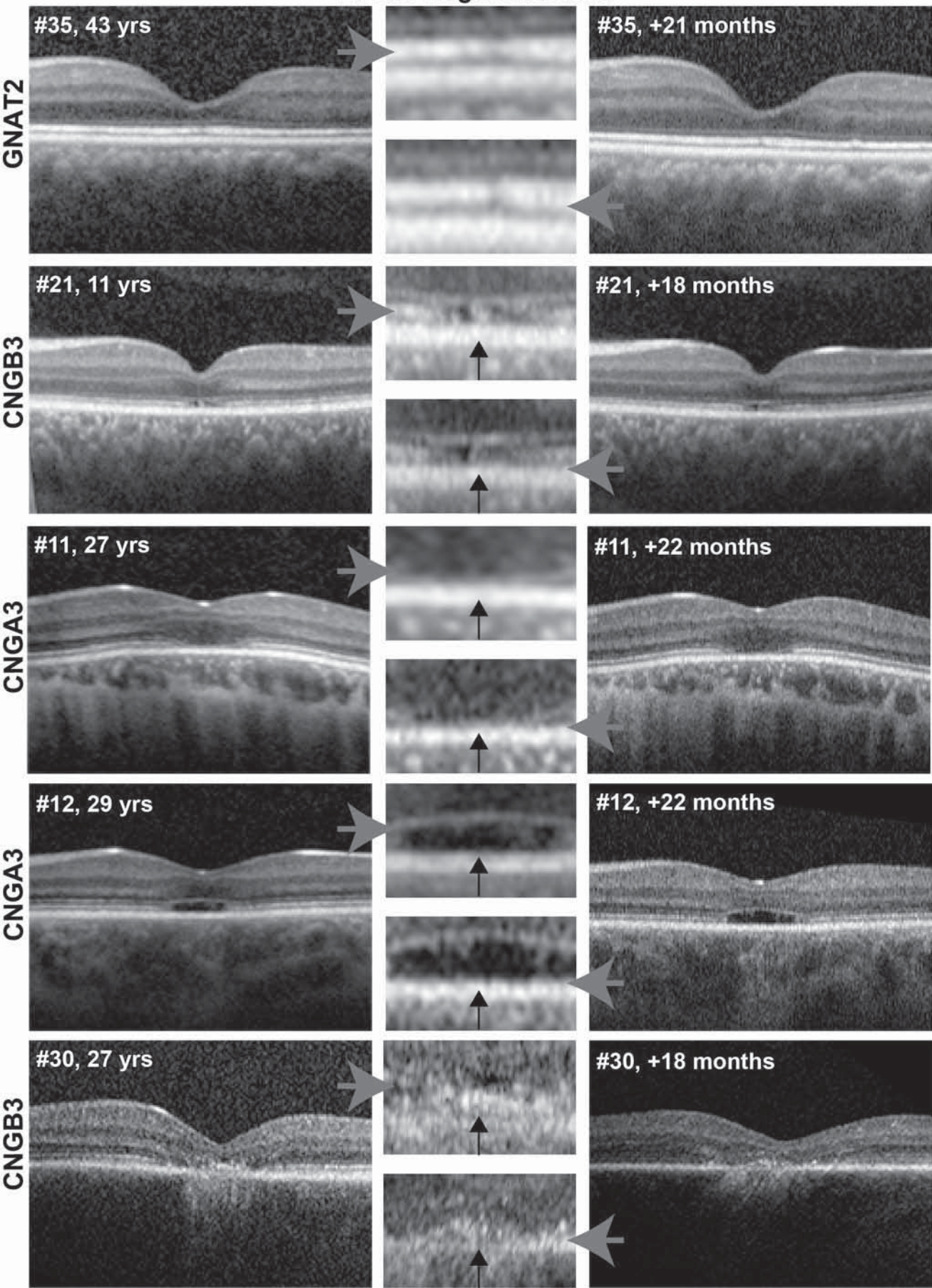
FIGURE 2 The left and right eye scans of the two achromatopsia subjects who demonstrated a change in SD-OCT appearance over the time course of the study. Layout is as in Figure 1. Top two panel rows: Patient #2 showed a continuous inner segment ellipsoid layer (ISe) at the fovea at initial scan in both the left (OS) and right (OD) eyes, and subsequently a disrupted ISe layer in both eyes at follow-up scan 20 months later (black arrows). Bottom two panel rows: Patient #31 showed a disrupted ISe layer at initial scan in both eyes and a hyporeflective zone (HRZ) 20 months later (black arrows). Non-magnified images represent 4500 μ m horizontally and 1000 μ m vertically; image scaling has been corrected for axial length. Scale bar, 200 μ m.

FIGURE 3 The microperimetry findings in patient #5 who developed a scotoma during this study. Left panel a): MP findings at baseline. Numbers indicate retinal sensitivity (dB) at that location. Right panel b): MP findings 16 months later indicate an absolute scotoma at two locations.

FIGURE 4 Longitudinal FAF in achromatopsia, demonstrating the three patterns observed (patient number, genotype and age at baseline acquisition indicated in left panels; patient number and follow-up interval in months indicated in right panels). Top panel row: Pattern 1: Reduced FAF signal centrally with a well-demarcated border. Middle panel row: Pattern 2: Normal FAF appearance.

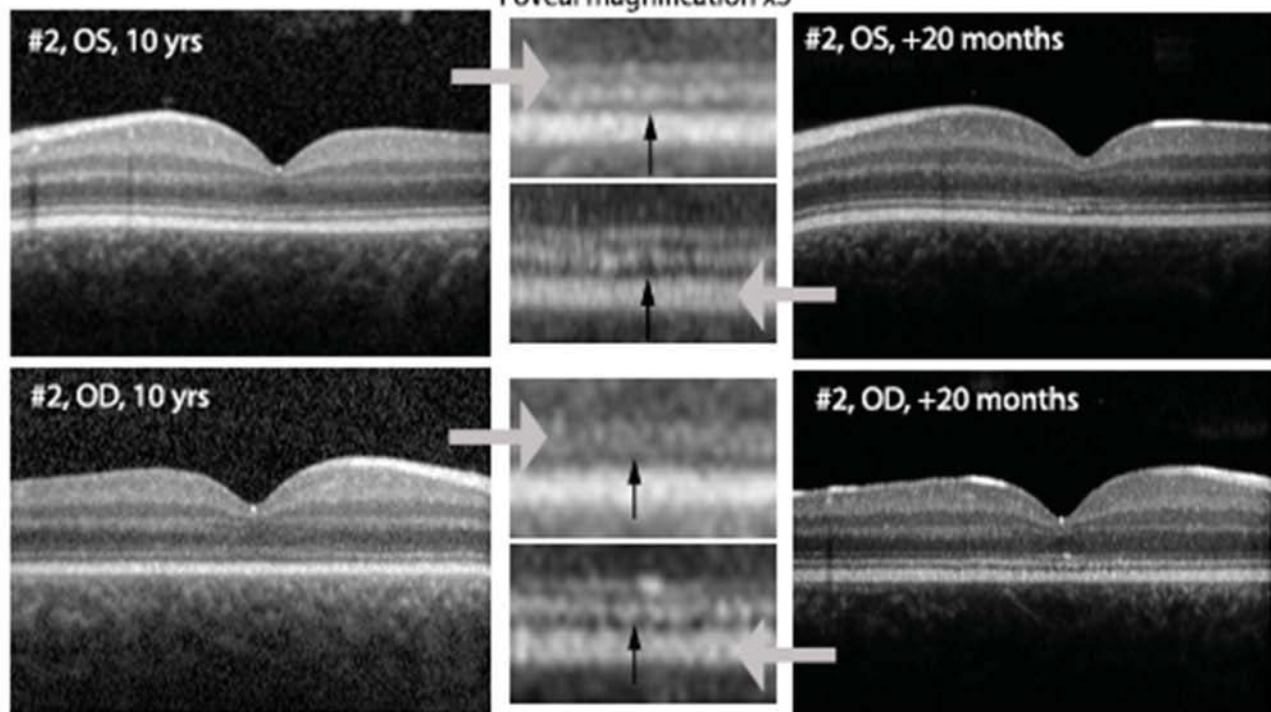
Bottom panel row: Pattern 3: A central increase in FAF. These patterns were not statistically significantly age dependent. All of the three above patients harbor *CNGA3* disease-causing alleles and are of a similar age, which serves to highlight both the wide phenotypic variation seen in achromatopsia even within the same genotype, and also the lack of strict age-dependency.

Foveal magnification x3

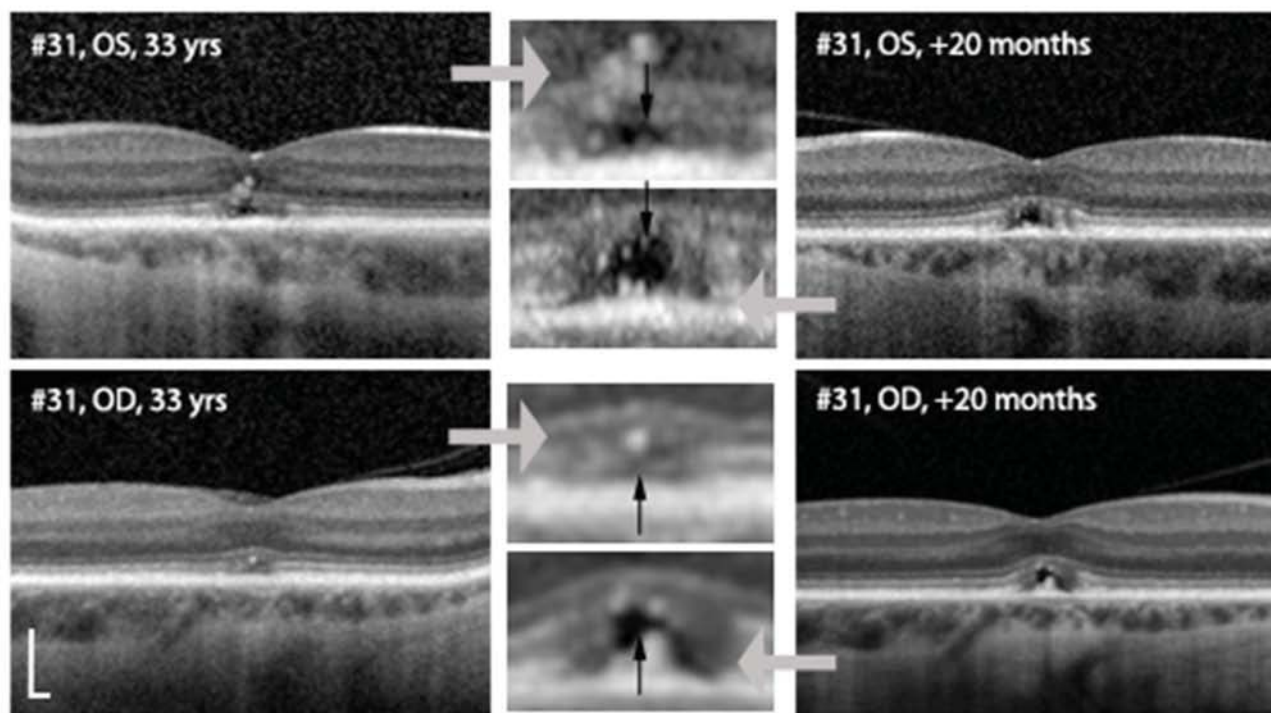


CNGA3

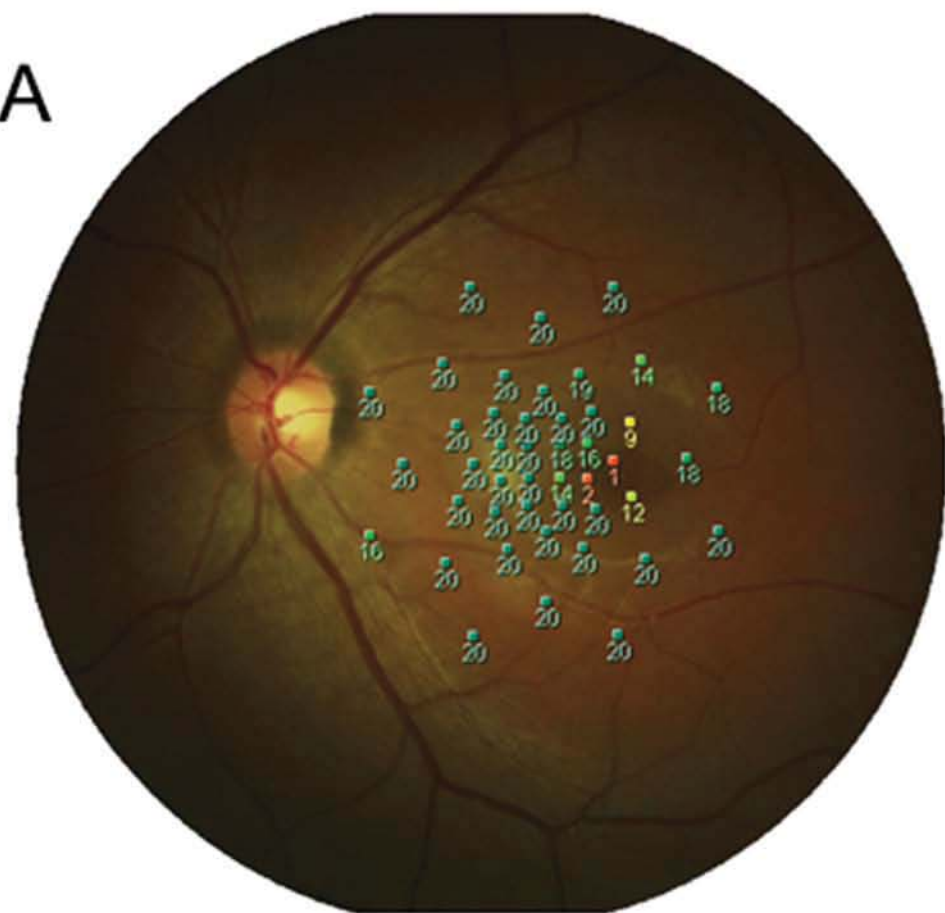
Foveal magnification x3



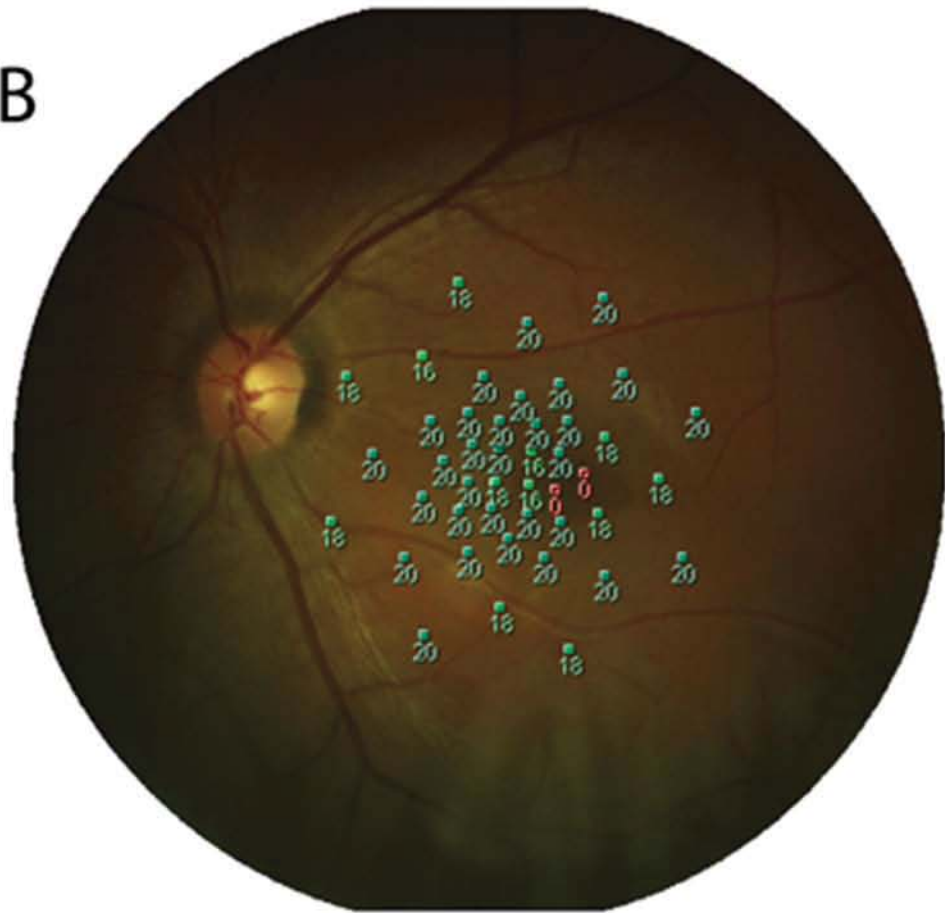
CNGB3



A



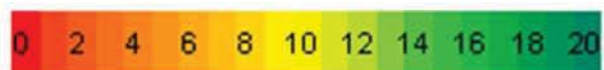
B



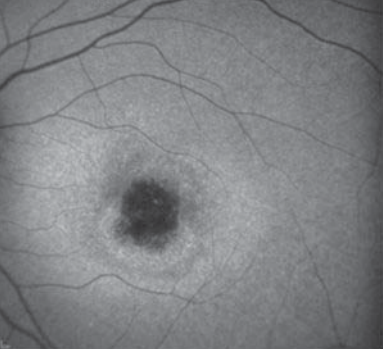
Legend

- Not seen at ... dB
- Seen at ... dB
- △ Not projected

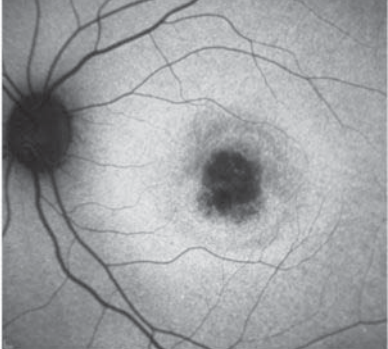
Attenuation scale (dB)



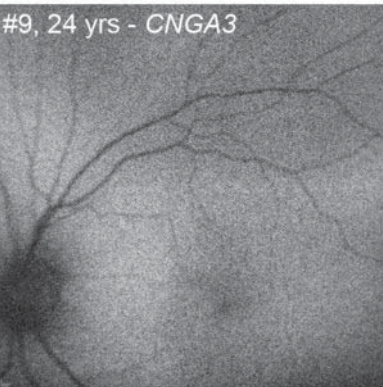
#10, 25 yrs - *CNGA3*



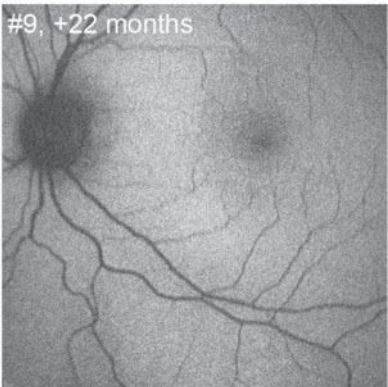
#10, +22 months



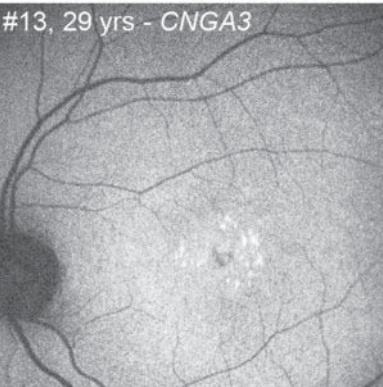
#9, 24 yrs - *CNGA3*



#9, +22 months



#13, 29 yrs - *CNGA3*



#13, +21 months



TABLE 1. Summary of Clinical Findings

Patient #	Age at baseline (yrs)	F/u interval (mnths)	Genotype	BCVA (logMAR)		Contrast sensitivity (logCS)		MP mean sensitivity (dB)		MP mean BCEA (°)		SD-OCT category*		Change in FTRT between assessments (µm)	Change in ONL thickness between assessments (µm)	Central FAF pattern at both assessments
				b	f/u	b	f/u	b	f/u	b	f/u	b	f/u			
1	7	21	CNGA3	0.94	0.86	1.30	1.40	14.90	16.20	44.04	36.70	3	N/A	N/A	N/A	N/A
2	10	20	CNGA3	0.80	0.82	1.25	1.55	19.67	19.75	15.82	8.05	1	2	+3	-10	Normal
3	11	16	CNGA3	1.08	1.04	0.90	1.10	14.90	14.50	40.70	61.80	1	1	+2	+2	N/A
4	11	17	CNGA3	0.94	1.00	1.05	1.30	17.60	19.50	14.75	17.90	2	2	+3	-1	Increased
5	17	16	CNGA3	0.80	0.86	1.20	1.35	18.35	18.55	6.00	7.35	3	3	-9	-1	Reduced
6	19	21	CNGA3	0.80	0.92	0.65	0.50	17.57	14.85	3.51	3.45	2	2	+2	0	Reduced
7	22	16	CNGA3	1.16	1.06	1.05	1.00	15.55	15.35	3.91	5.90	2	2	+3	+3	Normal
8	22	22	CNGA3	0.92	0.92	1.35	1.50	17.60	19.15	5.48	4.45	4	4	-3	-7	Normal
9	24	22	CNGA3	0.90	0.86	1.20	1.35	19.30	18.55	16.80	8.35	2	2	+2	+2	Normal
10	25	22	CNGA3	0.84	0.84	1.35	1.45	3.10	5.80	3.61	24.40	5	5	+4	N/A	Reduced
11	27	22	CNGA3	0.84	0.76	1.05	1.55	17.67	17.80	7.92	4.25	3	3	-7	-6	Reduced
12	29	22	CNGA3	0.74	0.90	1.20	1.25	13.55	15.20	37.43	7.60	4	4	+3	+2	Reduced
13	29	21	CNGA3	0.90	0.92	1.55	1.55	19.20	18.25	7.74	5.00	2	2	-8	-3	Increased
14	31	22	CNGA3	0.90	0.92	1.45	1.25	17.95	17.00	2.00	8.90	4	4	-2	+2	Normal
15	31	21	CNGA3	0.84	0.84	1.25	1.50	17.63	15.90	17.88	36.10	2	2	0	-4	N/A
16	34	21	CNGA3	0.90	0.82	1.20	1.05	13.80	14.90	10.20	14.40	2	2	-1	+3	Normal
17	35	21	CNGA3	0.96	0.96	0.90	1.00	14.17	15.90	13.15	7.60	2	2	-4	+2	Normal
18	49	17	CNGA3	0.80	0.74	1.30	1.30	17.15	18.25	2.23	2.95	4	4	+1	+2	Reduced
19	6	17	CNGB3	1.08	1.08	1.20	1.10	14.75	16.50	65.00	28.30	1	1	+2	-2	N/A
20	11	15	CNGB3	0.82	0.86	1.05	1.15	19.65	19.20	19.13	28.20	4	4	0	-1	Reduced
21	11	18	CNGB3	0.90	0.88	1.35	1.20	18.35	19.00	5.23	3.30	2	2	0	+3	Increased
22	13	21	CNGB3	0.90	1.00	1.00	1.10	18.93	19.25	8.62	8.80	3	3	-3	+3	Normal
23	12	16	CNGB3	0.76	0.80	1.30	1.35	19.80	19.05	5.97	3.95	2	2	+3	-4	Normal
24	13	18	CNGB3	0.90	0.82	1.30	1.50	19.27	19.95	6.54	6.00	1	1	-8	+1	Increased
26	18	24	CNGB3	0.74	0.76	1.25	1.40	18.40	17.80	3.70	4.10	4	4	+3	+2	Reduced
27	19	17	CNGB3	0.86	0.88	1.35	1.35	18.50	17.10	3.75	2.60	4	4	+3	-3	N/A
28	23	13	CNGB3	0.76	0.80	1.25	1.55	18.35	17.75	6.32	6.40	5	5	+4	N/A	Reduced
29	24	20	CNGB3	0.90	0.92	1.45	1.65	15.00	14.80	3.27	3.60	1	1	-2	-2	N/A
30	27	18	CNGB3	1.20	1.04	1.15	1.00	14.63	15.75	38.02	44.95	5	5	+2	N/A	Reduced
31	33	20	CNGB3	0.96	0.84	0.90	1.20	16.65	18.00	15.12	11.90	2	4	+3	+3	Reduced
33	47	13	CNGB3	0.96	1.00	1.10	1.15	19.35	17.00	7.30	5.00	4	4	+3	-1	Reduced
34	29	24	GNAT2	0.88	0.94	1.20	0.25	14.30	14.80	2.09	1.70	4	4	-2	0	Reduced
35	43	21	GNAT2	1.10	1.10	0.70	0.75	14.83	12.30	16.56	15.70	1	1	-4	-1	Reduced
36	49	21	GNAT2	0.94	1.04	1.05	0.75	13.43	13.20	10.31	11.70	1	1	-1	-1	Normal
37	52	21	GNAT2	1.00	1.00	0.50	0.65	14.80	14.30	20.90	18.95	1	1	+1	0	Normal
38	43	21	PDE6C	1.32	1.28	1.05	0.80	7.77	7.70	17.15	11.85	3	3	+2	+1	Reduced
39	19	21	?	0.96	0.92	1.45	1.55	19.57	18.80	9.54	20.10	3	3	+2	-5	N/A
40	23	21	?	0.86	0.74	1.20	1.35	19.83	19.75	16.14	10.85	3	3	0	0	N/A

#, patient number (as in Sundaram et al¹⁹); b, baseline; f/u, follow-up; BCVA, best corrected visual acuity; OD, right eye; OS, left eye; LogMAR, logarithm of the minimum angle of resolution; LogCS, logarithm of contrast sensitivity; MP, microperimetry; dB, decibels; BCEA, bivariate contour ellipse area; SD-OCT, spectral domain optical coherence tomography; ISe, inner segment ellipsoid; RPE, retinal pigment epithelium; N/A, not assessable. FTRT, foveal total retinal thickness. ONL, outer nuclear layer. '+' indicates an increase in metric over time between assessments; '-' indicates a reduction over time; FAF, fundus autofluorescence; '?', no disease causing variant identified in *CNGA3*, *CNGB3*, *GNAT2*, *PDE6C* or *PDE6H*. *SD-OCT category: 1 = continuous ISe; 2 = ISe disruption; 3 = ISe absence; 4 = hyporeflexive zone present; 5 = outer retinal atrophy.